**Editorial comments:**  
  
1. There are still many commercial terms in the manuscript (in particular mTeSR1, AggreWell, Matrigel, and Accutase). To avoid the appearance of being an advertisement, these must be reduced to ideally less than 10 instances in total; also, none should be in the abstract, keywords, or protocol headings. I do understand that this protocol relies on particular reagents and equipment-can you, say, introduce more generic terms the first time you mention particular products?

>> We removed AggreWell from the keyword list. We also removed mTeSR1, Matrigel, Accutase, and AggreWell from the long abstract.

A reviewer of the original manuscript requested us to state that mTeSR1 is “a ready-to-use, commercially available medium formulated for maintenance of human PSCs in the primed pluripotency state.” In the R1 we included this statement in the long abstract. In the R2, we moved it to the first mention to mTeSR1 in the introduction.

We introduced more generic terms at the first mention to Matrigel in the introduction – namely, “… dishes coated with the Matrigel extracellular matrix protein.”

We introduced more generic terms at the first mention to Accutase (1.3.1) – namely, “Accutase cell dissociation enzyme mixture.”

In the protocol text of the R2, we attempted to reduce mentions to the commercial names of the reagents to the extent that the lack of commercial names does not introduce ambiguity.

2. The abstract, introduction, and discussion all mention the importance of the short incubation time in mTeSR1 and 4i, but you only have chosen to film the latter part of this procedure. There is still room to at least briefly show the earlier steps.

>> The R2 manuscript added 2.2.9, 2.2.10, 2.3.2, 2.4.3.4, 2.4.3.8, and 2.4.4.6 for filming.

3. 2.5.2.6 is still a little unclear-how exactly are you adding hpGCLC medium at the end? Through the membrane?

>> We revised the last part of 2.5.2.6 as follows:

Collect EBs in the conical tube by adding 18 mL prewarmed hPGCLC complete medium from above the membrane of cell strainer. Thus, medium will go through the membrane and collect EBs attached on the lower side of the membrane down to the bottom of the centrifuge tube.

4. Figure 9 legend: What does ‘EB, non-hPGCLC cells’ mean? Also, it seems fairly clear, but could you indicate what the colors means?

>> We revised Fig 9 legend as follows:

hPGCLCs can be enriched as CD38+ cells by FACS (red dots). EB cells that do not express CD38 (blue dots) should also be collected as negative control. FACS gates of CD38-positive and CD38-negative cells should be separated with a wide margin (green dots) to avoid contamination of each type of cells.

5. Discussion: Can you explain a bit further what you mean by a ‘rhBMP4 lot check’; possibly with references?

>> Because this is our previously undisclosed technical tip, there is no citable reference. We revised this part of the discussion as follows:

We recommend to test several different lots of recombinant human BMP4 reagents obtained from multiple vendors for their performance in supporting hPGCLC generation and secure a large amount of the best lot.